



## VALUE OF SERUM INTERLEUKIN-22 IN PATIENTS WITH CHRONIC LYMPHOCYTIC LEUKEMIA AND ITS RELATION TO DISEASE CHARACTERISTICS.

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### ABSTRACT:

**Background:** Chronic lymphocytic leukaemia (CLL) is a common type of leukaemia characterized by progressive accumulation of monoclonal B cells. The survival and proliferation of CLL cells are stimulated by T-helper cytokines and could be correlated with disease progression. The aim of this study is to assess the value of serum interleukin-22 in patients with chronic lymphocytic leukemia and its relation to disease characteristics.

**Methods:** The study was carried out on 40 CLL patients (newly diagnosed) and 10 healthy controls. Routine hematological and laboratory investigations were performed for all subjects in addition to interleukin-22 (IL-22) level and ZAP-70 expression.

**Results:** Mean values of IL-22 level among CLL patients were significantly higher than in the control group. IL-22 mean values were significantly higher in patients with positive ZAP-70 expression. Moreover, there was a significant positive correlation between serum IL-22 level and ZAP-70. According to the ROC curve, the area under the curve was statistically significant (AUC=0.809 p=0.001).

**Conclusion:** IL-22 & ZAP-70 could act in alliance to maintain survival of CLL cells and its proliferation and could be used as diagnostic as well as prognostic markers. Our findings strongly recommend the incorporation of IL-22 level into routine CLL work up.

**Keywords:** Chronic lymphocytic leukemia, ZAP-70, Interleukin-22 (IL-22),  $\beta$ 2-microglobulin, LDH

### 1. INTRODUCTION

Chronic lymphocytic leukemia (CLL) is defined as a low-grade lymphoproliferative neoplasm with  $\geq 5 \times 10^9/L$  clonal B-cells in the peripheral circulation which express CD5, CD19, CD20, and CD23 by the World Health Organization and the International Consensus Classification Clinical Advisory Committee [1, 2]. For evaluation of CLL patients, Rai and Binet are the two staging systems commonly used [3]. They depend on physical evidence (ie, lymphadenopathy,

splenomegaly and/or hepatomegaly) and blood parameters (anemia or thrombocytopenia) to assess the degree of tumor burden and predict treatment requirements and survival. However, it cannot determine patients who may develop aggressive disease [4, 5].

ZAP-70 (70-kDa zeta-associated protein) is an intracellular tyrosine kinase that has a major role in T-cell signaling. It is also associated with the B-cell receptor in CLL. ZAP-70 expression ( $\geq 20\%$  of B cells) is

associated with an increased risk for the development of adverse outcomes in B-CLL. Thus, it is considered a prognostic factor for these patients [6]. Previous studies reported a correlation between ZAP-70 expression in CLL cells and Immunoglobulin Heavy Chain Gene Mutation (IGHV) status. ZAP-70 positive patients have an aggressive course, an immediate treatment requirement, and lower survival rates. If ZAP-70 expression is present, it remains constant all over the clinical course of the patient, and consequently, it is a valid risk marker regardless of the evaluation's timing [7, 8].

IL-22 is a cytokine that plays a crucial role in many physiological processes, ranging from innate immune responses to tissue regeneration. When its activity is impaired, this can lead to chronic inflammation, impaired wound healing, and infections [3, 4]. IL-22 has tumor-promoting properties such as tumor-cell proliferation, anti-apoptosis, and attraction of immunosuppressive immune cells [9], as well as promoting neoangiogenesis and transition from epithelial to mesenchymal, which are hallmarks of cancer [10]. IL-22 is linked to the pathogenesis of cancer colon in both inflammatory and genetic models [11] and also is linked to more aggressive phenotypes in lung, breast, gastric, and skin cancers, indicating the role of IL-22 in cancer progression [12, 13]. IL-22 up-regulation and down-regulation generate many consequences that determine its biological and pathological activities [14, 15]. Its dual role highlights the therapeutic potential of adjusting the cytokine network to

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**Table 2: ZAP-70 expression in relation to staging of CLL patients**

Parameter	Zap-70 expression				p-value <sup>a</sup>
	ZAP-70 positive (n=26) (65%)		ZAP-70 negative (n=14) (35%)		
	N <sub>o</sub>	%	N <sub>o</sub>	%	
<b>- Rai staging:</b>					
I (n= 1) (2.5%)	0	0.0	1	7.1	<b>0.002*</b>
II (n= 10) (25.0%)	2	7.7	8	57.1	
III (n= 18) (45.0%)	15	57.7	3	21.4	
IV (n= 11) (27.5%)	9	34.6	2	14.3	
<b>- Binet staging:</b>					
A (n= 7) (17.5%)	3	11.5	4	28.6	0.235
B (n= 12) (30.0%)	7	26.9	5	35.7	
C (n= 21) (52.5%)	16	61.5	5	35.7	

\*p-values are statistically significant

<sup>a</sup>Chi-square test

#### Serum IL-22

Mean serum concentration of IL-22 showed significantly higher values among CLL patients compared to the control group (47.01 vs. 10.64 pg/ml) (p =0.0001). Table 3 shows the

mean values of serum IL-22 according to the staging of CLL patients. There were significant differences in mean serum IL-22 regarding Rai and Binet staging of CLL patients (p = 0.0001,p=0.018,respectively).

**Table 3: Serum IL-22 according to staging of CLL patients**

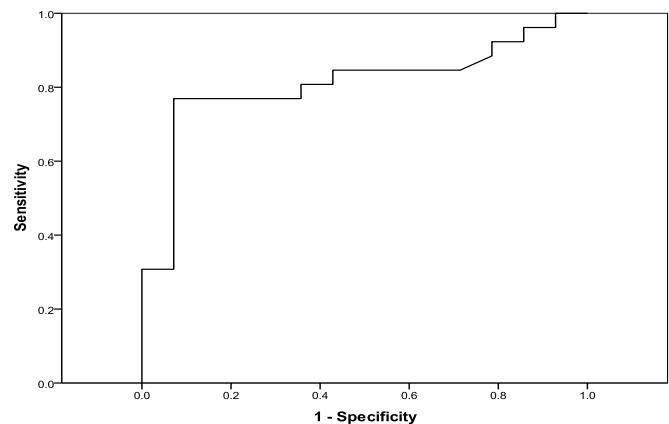
Parameter	IL-22 (pg/ml)		p-value <sup>a</sup>
	Median	Mean ± SD (Min. – Max.)	
<b>- Rai staging:</b>			
I (n= 1)	31.59	31.59	<b>0.0001*</b>
II (n= 10)	37.19	37.36 ± 8.75 (26.68 - 54.42)	
III (n= 18)	46.85	44.85 ± 11.90 (20.68 - 62.09)	
IV (n= 11)	59.70	60.70 ± 10.37 (42.87 - 75.83)	
<b>- Binet staging:</b>			
A (n= 7)	31.59	38.68 ± 11.78 (26.68 - 54.42)	<b>0.018*</b>
B (n= 12)	38.86	41.96 ± 10.54 (28.14 - 62.09)	
C (n= 21)	53.87	52.66 ± 14.08 (20.68 - 75.83)	

\*p-values are statistically significant  
SD: standard deviation

<sup>a</sup>Kruskal-Wallis test

**IL-22 & ZAP-70**

Significantly higher mean values of serum IL-22 were observed among ZAP-70 positive compared to ZAP-70 negative patients (51.94 vs. 37.83 pg/ml) ( $p = 0.001^*$ ). The IL-22 ideal cut-off value was established at 41.22 by the ROC curve to evaluate IL-22 as a sensitive predictor of ZAP-70 positivity. In addition, 84.62% was the sensitivity, 71.43% was the specificity, PPV was 84.6%, and NPV was 71.4%. The area under the curve was statistically significant (AUC = 0.809, 95% CI=0.669 - 0.949,  $p = 0.001$ ). (Figure 1) (Table 4)



Diagonal segments are produced by ties.

**Figure1: ROC curve for detection of IL-22 as a sensitive predictor of ZAP-70 positivity**

**Table 4: Prognostic performance for detection of IL-22 as a sensitive predictor of ZAP-70 positivity**

	AUC	p	95% C.I	Cut off#	Sensitivity	Specificity	PPV	NPV
<b>IL – 22</b>	0.809*	0.001*	0.669 – 0.949	> <b>41.22</b>	84.62	71.43	84.6	71.4

AUC: Area Under a Curve

p value: Probability value

CI: Confidence Intervals

NPV: Negative predictive value

PPV: Positive predictive value

\*: Statistically significant at  $p \leq 0.05$

IL-22 level showed a significant positive correlation to  $\beta$ 2-microglobulin ( $r=0.448$ ), LDH ( $r=0.463$ ), CRP ( $r= 0.462$ ) and ZAP-70 ( $r= 0.459$ ), while other hematological parameters and

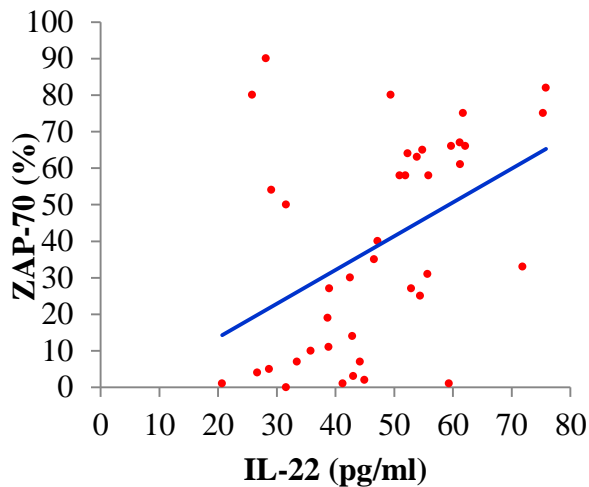
serum markers did not show significant correlation. (Table 5, Figure2).

**Table 5a: Correlation of hematological and biochemical markers to IL-22 in CLL patients**

Variables	IL-22 (pg/ml)	
	r	P
Hemoglobin concentration (g/dl)	- 0.307	0.054
Total leukocytic count ( $\times 10^9/L$ )	- 0.156	0.337
Absolute lymphocytes count ( $\times 10^9/L$ )	- 0.166	0.305
Platelets count ( $\times 10^9/L$ )	- 0.227	0.159
Reticulocytes count (%)	0.078	0.632
$\beta$ 2-microglobulin (mg/L)	0.448*	0.004*
Lactate dehydrogenase (U/L)	0.463*	0.003*
C-reactive protein (mg/dl)	0.462*	0.003*
SGPT(ALT) (U/L)	- 0.051	0.756
SGOT(AST) (U/L)	0.094	0.564
Serum urea (mg/dl)	- 0.167	0.302
Serum creatinine (mg/dl)	- 0.055	0.738
ZAP-70 expression (%)	0.459*	0.003*

r: Spearman correlation coefficient

\* Significant statistical correlation



**Figure 2: Positive correlation between IL-22 and ZAP-70**

### Discussion

CLL is characterized by clonal proliferation and accumulation of mature CD5-positive B-cells in blood, bone marrow, lymph nodes, and spleen. Apparently, the development of clonal B cells occurs at the hematopoietic stem cell (HSC) stage, raising the possibility that multipotent HSCs are involved in the primary leukemogenic state in CLL [5].

The diagnosis of CLL requires the presence of  $\geq 5000$  B-lymphocytes/ $\mu\text{L}$  in the peripheral blood (not less than 3 months). The clonality of the circulating B-lymphocytes is confirmed by flow cytometry. The blood smears of CLL patients show mature lymphocytes that are small in size with a narrow border of cytoplasm and a dense nucleus lacking discernible nucleoli and having partially aggregated chromatin [29].

Clinical progression of CLL patients varies largely, as some patients show rapid progression while others don't require treatment and live for years. Most patients are diagnosed in early stages; thus, Rai and Binet staging systems do not identify patients with poor prognosis [30]. Immunoglobulin heavy-chain gene (IGHV) mutation status, cytogenetic abnormalities, and expression of certain proteins have emerged as important prognostic indicators for CLL. For many clinical laboratories, routine analysis is difficult and expensive [31]. As a result, readily accessible surrogate markers are needed.

ZAP-70 expression in CLL is linked to IGHV mutational status, survival, and disease progression. Previous studies demonstrated that aberrant B-cell receptor (BCR) signalling, proliferation, and migration towards tumor microenvironment are all influenced by the abnormal expression of the ZAP-70 protein in CLL cells [30, 32].

In the current study, 65.0% of CLL patients have positive ZAP-70 expression. This was in accordance with the percentage reported by Singleterry et al. [33], but Zeeshan et al. [34] found that ZAP-70 was positive in 12 (13.5%) patients, while 77 (86.5%) were negative. Previous studies by Liu et al. [30] concluded that ZAP-70 is overexpressed in

IGHV-unmutated CLL but not in normal "B" lymphocyte cells and showed a similar clinical value to IGHV mutational status in terms of disease progression and survival.

In the present work, ZAP-70 expression showed a highly significant association with Rai staging ( $p = 0.002$ ) but not with Binet staging. This finding was in accordance with the study of Patel [35], who stated that in Binet staging, there was no significant difference in ZAP-70 expression. Zeeshan R et al. [34] revealed that ZAP-70 protein and Rai stage III disease were found to have a high positive correlation ( $p=0.004$ ). These factors imply that ZAP-70 expression in CLL may represent a malignant clone in an activated state and be linked to a progressing disease. As a result, a laboratory test for ZAP-70 expression may be a crucial addition to the patient's overall care.

In the present study, patients with positive ZAP-70 expression had a significantly higher median level of  $\beta 2$ -microglobulin compared to negative ZAP-70-expressing patients ( $p = 0.011$ ). Serum LDH was also higher in positive ZAP-70 than in negative-expressing patients but was not statistically significant. The study of Assem et al. [36] reported a significant rise in LDH and  $\beta 2\text{M}$  serum levels in ZAP-70 positive groups when compared to negative groups and concluded that ZAP-70 is predictive of disease progression. Amaya-Chanaga et al. [37] declared that, while choosing the ideal plan of treatment and prognosis for CLL patients, physicians must take ZAP-70 expression into account as one of the independent prognostic tools.

IL-22 is a glycoprotein released by several types of CD4-positive cells, including Th-22 cells. According to reports, it contributes to the development and progression of malignancies. The IL-22/IL-22 receptor-1 signalling cascade is another route by which cancer stem cells can endure and multiply. Additionally, it is clear that IL-22 levels are elevated in many types of cancers [38].

The mean level of IL-22 in our study was significantly higher in CLL patients than in the control group. This is in concordance with Heiba et al. [38] and Abd El-Hamed et al. [39], who revealed an elevation in IL-22 plasma levels in CLL patients compared to healthy subjects. Lim et al. [10] and Voigta et al. [9] reported that IL-22 is a cancer-promoting cytokine.

Our work demonstrates a highly significant difference between the level of serum IL-22 according to Rai staging ( $p = 0.0001$ ) and Binet staging of CLL patients ( $p = 0.018$ ). However, in contrast to our results, Gangemi et al. [40] reported that throughout the various stages of CLL, there were no statistically significant changes in the levels of IL-22. Also, Kouzegaran et al. [41] found that levels of IL-22 were not significantly associated with the different stages of disease (Rai stages). Our results were partly concordant with those of Atreya et al. [42], who showed a direct association between staging of colorectal cancer and IL-22, which confirms the involvement of IL-22 in the progression of carcinogenesis.

Our results indicate that CLL patients presenting with positive expression of ZAP-70 had significantly higher IL-22 levels in comparison with those negative ZAP-70. This finding is in accordance with Heiba et al. [38], who found

that patients with positive ZAP-70 expression had higher plasma levels of IL-22 than healthy individuals. Abd El-Hamed et al. [39] suggested that a high level of IL-22 and positive ZAP-70 expression have a synergistic action in inhibiting apoptosis and triggering proliferative responses. Finally, this study found a highly significant correlation between IL-22 and  $\beta$ 2-microglobulin, LDH, CRP, and ZAP-70, while other hematological parameters and serum markers did not show a statistically significant correlation. In accordance with our result, Kouzegaran et al. [41] found that CLL patients with positive CD38 and ZAP-70 had higher plasma levels of IL-22 as compared to healthy individuals. On the contrary, Gangemi et al. [43] reported that levels of ZAP-70, LDH, and  $\beta$ 2-microglobulin did not significantly correlate with plasma IL-22 levels. Protopsaltis et al. [44] demonstrated that IL-22 can induce angiogenesis by directly acting on endothelial cells. Blockage of IL-22 inhibits tumor growth and reduces tumor angiogenesis, thus providing a possible cancer treatment through the development of anti-IL-22 therapies.

### Conclusion

According to these findings, which denote the sensitivity of serum IL-22 in predicting ZAP-70 positivity among CLL patients, serum IL-22 could be used as a surrogate and reliable sole marker for ZAP-70 expression. In addition, IL-22 may be used as a novel goal for CLL immunotherapy approaches.

### Recommendations

Assessment of IL-22 should be included in the CLL work-up.

### References

- Alaggio R, Amador C, Anagnostopoulos I, Attygalle AD, Araujo IBO, Berti E, Bhagat G, Borges AM, Boyer D, Calaminici M et al: The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Lymphoid Neoplasms. *Leukemia* 2022, 36(7):1720-1748.
- Campo E, Jaffe ES, Cook JR, Quintanilla-Martinez L, Swerdlow SH, Anderson KC, et al. The International Consensus Classification of Mature Lymphoid Neoplasms: a report from the Clinical Advisory Committee. *Blood* 2022; 140:1229–53.
- Slager SL, Parikh SA, Achenbach SJ, Norman AD, Rabe KG, Boddicker NJ, et al. Progression and survival of monoclonal B-cell lymphocytosis (MBL): a screening study of 10,139 individuals. *Blood*. 2022; 140:1702–9.
- Hampel PJ, Parikh SA. Chronic lymphocytic leukemia treatment algorithm 2022. *Blood Cancer J*. 2022 Nov 29; 12(11):161. doi: 10.1038/s41408-022-00756-9. Erratum in: *Blood Cancer J*. 2022 Dec 22; 12(12):172.
- Hallek M, Al-Sawaf O. Chronic lymphocytic leukemia: 2022 update on diagnostic and therapeutic procedures. *Am J Hematol*. 2021 Dec 1; 96 (12):1679-1705. doi: 10.1002/ajh.26367.
- Isiksacan N, Cinar S, Aktas Cetin E, Aktan M, Deniz G. Cytokine Contents in Chronic Lymphocytic Leukemia: Association with ZAP70 Expression. *Turk J Haematol*. 2016; 33(3):202-8.
- Wiggers TG, Westra G, Westers TM, Abbes AP, Strunk A, Kuiper-Kramer E, et al. ZAP70 in B-CLL cells related to the expression in NK cells is a surrogate marker for mutational status. *Cytometry B Clin Cytom*. 2014; 86(4):280-7.
- Preobrazhensky SN, Szankasi P, Bahler DW. Improved flow cytometric detection of ZAP-70 in chronic lymphocytic leukemia using experimentally optimized isotypic control antibodies. *Cytometry B Clin Cytom*. 2012; 82(2):78-84.
- Voigt C, May P, Gottschlich A, Markota A, Wenk D, Gerlach I, et al. Cancer cells induce interleukin-22 production from memory CD4+ T cells via interleukin-1 to promote tumor growth. *Proc Natl Acad Sci U S A*. 2017;114(49):12994-12999.doi: 10.1073/pnas.1705165114.
- Lim C, Savan R. The role of the IL-22/IL-22R1 axis in cancer. *Cytokine Growth Factor Rev*. 2014 Jun;25(3):257-71. doi: 10.1016/j.cytogfr.2014.04.005.
- Kryczek I, Lin Y, Nagarsheth N, Peng D, Zhao L, Zhao E, et al. IL-22(+) CD4 (+) T cells promote colorectal cancer stemness via STAT3 transcription factor activation and induction of the methyltransferase DOT1L. *Immunity*. 2014;40(5):772-784. doi: 10.1016/j.immuni.2014.03.010.
- Kim K, Kim G, Kim JY, Yun HJ, Lim SC, Choi HS. Interleukin-22 promotes epithelial cell transformation and breast tumorigenesis via MAP3K8 activation. *Carcinogenesis*. 2014 Jun;35(6):1352-61. doi: 10.1093/carcin/bgu044.
- Kobold S, Völk S, Clauditz T, Küpper NJ, Minner S, Tufman A, et al. Interleukin-22 is frequently expressed in small- and large-cell lung cancer and promotes growth in chemotherapy-resistant cancer cells. *J Thorac Oncol*. 2013Aug;8(8):1032-42.doi: 10.1097/JTO.0b013e31829923c8.
- Tanzeela Arshad, Fizzah Mansur, Richard Palek, Sobia Manzoor, and Vaclav Liska .A double edged sword role of interleukin -22 in wound healing and tissue regeneration.*Frontiers in Immunology* 2020;11:2148.
- Lanfranca MP, Lin Y, Fang J, Zou W, Frankel T. Biological and pathological activities of interleukin-22. *J Mol Med (Berl)*. 2016;94(5):523-34. doi: 10.1007/s00109-016-1391-6.
- Shabgah AG, Navashenaq JG, Shabgah OG, Mohammadi H, Sahebkar A: Interleukin-22 in human inflammatory diseases and viral infections. *Autoimmunity reviews* 2017, 16(12):1209-1218.
- Hernandez P, Gronke K, Diefenbach A. A catch-22: Interleukin-22 and cancer. *Eur J Immunol*. 2018;48(1):15-31. doi: 10.1002/eji.201747183.
- Hallek M, Cheson BD, Catovsky D, Caligaris-Cappio F, Dighiero G, Döhner H, et al. International Workshop on Chronic Lymphocytic Leukemia. Guidelines for the diagnosis and treatment of chronic lymphocytic leukemia: a report from the International Workshop on Chronic Lymphocytic Leukemia updating the National Cancer Institute-Working Group 1996 guidelines. *Blood*.

- 2008; 111 (12):5446-56. doi: 10.1182/blood-2007-06-093906.
19. Wierda WG, Brown J, Abramson JS, Awan F, Bilgrami SF, Bociek G, et al: NCCN Guidelines® Insights: Chronic Lymphocytic Leukemia/Small Lymphocytic Lymphoma, Version 3.2022. Journal of the National Comprehensive Cancer Network: JNCCN 2022, 20(6):622-634.
  20. Bain BJ, Bates I, Laffan MA, and Lewis SM: Basic hematological techniques, Dacie and Lewis practical haematology, 12th edition.p.19-47 London: Churchill Livingstone; 2017.
  21. Bates I., Burthem J. Bone marrow biopsy. Dacie and Lewis practical haematology, 12th edition.p.112-125 London: Churchill Livingstone; 2017.
  22. Morilla AM., Morilla RR., Nadal-Melsio E. Immunophenotyping by flow cytometry. Dacie and Lewis practical haematology, 12th edition.p.330-349 London: Churchill Livingstone; 2017.
  23. Panteghini M. Serum enzymes. Tietz fundamentals of clinical chemistry (7th ed p. 318-336). St Louis, Missouri: Elsevier, Co, 2015
  24. Hortin GL. Amino acids, peptides, and proteins. Tietz fundamentals of clinical chemistry (7th ed p. 318-336). St Louis, Missouri: Elsevier, Co., 2015
  25. Janelle M, Xin XU. Liver function. In M. L. Bishop, E. P. Fody & L. E. Schoeff (Eds.), Clinical chemistry principles, procedures and correlations (8th edition p.1266-1324). Lippincott Williams and Wilkins, 2018
  26. Lamb EJ, & Price CP. Kidney function tests- creatinine, urea, and uric acid, Tietz fundamentals of clinical chemistry (7th ed p. 364-375). St Louis, Missouri: Elsevier, Co., 2015
  27. Preobrazhensky SN, Bahler DW. Optimization of flow cytometric measurement of ZAP-70 in chronic lymphocytic leukemia. Cytometry B Clin Cytom. 2008; 74(2):118-27.
  28. Ruggeri RM, Minciullo P, Saitta S, Giovinazzo S, Certo R, Campenni A, et al. Serum interleukin-22 (IL-22) is increased in the early stage of Hashimoto's thyroiditis compared to non-autoimmune thyroid disease and healthy controls. Hormones (Athens). 2014; 13(3):338-44.
  29. Hallek M, Cheson BD, Catovsky D, Caligaris-Cappio F, Dighiero G, Döhner H, et al. iwCLL guidelines for diagnosis, indications for treatment, response assessment, and supportive management of CLL. Blood. 2018;131(25):2745-2760. doi: 10.1182/blood-2017-09-806398.
  30. Liu Y, Wang Y, Yang J, Bi Y, Wang H. ZAP-70 in chronic lymphocytic leukemia: A meta-analysis. Clin Chim Acta. 2018;483:82-88. doi: 10.1016/j.cca.2018.04.026.
  31. Parikh SA, Shanafelt TD: Prognostic factors and risk stratification in chronic lymphocytic leukemia. Seminars in Oncology 2016, 43(2):233-240.
  32. Wagner M, Oelsner M, Moore A, Götte F, Kuhn PH, Haferlach T, et al. Integration of innate into adaptive immune responses in ZAP-70-positive chronic lymphocytic leukemia. Blood. 2016, 127(4):436-448.
  33. Singletary WL, Cruse JM, Lewis RE, Coker JA, Suggs JL. CD49d, Zap-70, and CD38 Expression in Chronic Lymphocytic Leukemia. FASEB J. 2009;23:740-5
  34. Zeeshan R, Irfan SM, Sultan S, Bhimani S. ZAP-70 protein expression in B-cell chronic lymphoid leukemia: a single center experience from Pakistan. Asian Pac J Cancer Prev. 2015; 16 (4):1587-90. doi:10.7314/APJCP.2015.16.4.1587
  35. Patel DY. Comparison of  $\beta$ -2 microglobulin with CD38 and zap-70 as prognostic markers in chronic lymphocytic leukemia [M.D. Thesis submitted to Human Pathology Department]: University of Nairobi; 2013. pages 105.
  36. Assem M, Abdel Hamid T, Kohla S, Arsanyos S. The Prognostic Significance of Combined Expression of ZAP-70 and CD38 in Chronic Lymphocytic Leukemia. J Egypt Natl Canc Inst. 2009; 21(4):287-97.
  37. Amaya-Chanaga CI, Rassenti LZ. Biomarkers in chronic lymphocytic leukemia: Clinical applications and prognostic markers. Best Pract Res Clin Haematol. 2016; 29(1):79-89. doi: 10.1016/j.beha.2016.08.005.
  38. Heiba N, Elshazly S: Plasma interleukin-22 and its cellular receptor (IL-22RA1) expression in chronic lymphocytic leukemia. The Egyptian Journal of Haematology. 2013, 38(4):123-129.
  39. Abd El-Hamed NA, El Sharkawi EA, Abd El-Naiem EA, Abd El-Hamed WM. Significance of Interleukin-22 and CD38 in Chronic Lymphocytic Leukemia. MJMR, 2019;30(4), 365-371
  40. Gangemi S, Allegra A, Alonci A, Pace E, Ferraro M, Cannavo A, et al. Interleukin 22 is increased and correlated with CD38 expression in patients with B-chronic lymphocytic leukemia. Blood Cells Mol Dis. 2013; 50(1):39-40.
  41. Kouzegaran S, Siroosbakht S, Farsad BF, Rezakhaniha B, Dormanesh B, Behnod V, et al. Elevated IL-17A and IL-22 regulate expression of inducible CD38 and Zap-70 in chronic lymphocytic leukemia. Cytometry B Clin Cytom. 2018; 94(1):143-147. doi: 10.1002/cyto.b.21487.
  42. Atreya I, Kindermann M, Wirtz S. Innate lymphoid cells in intestinal cancer development. Semin Immunol. 2019;41:101267. doi: 10.1016/j.smim.2019.02.001.
  43. Gangemi S, Allegra A, Alonci A, Pace E, Ferraro M, Cannavo A, et al. Interleukin 22 is increased and correlated with CD38 expression in patients with B-chronic lymphocytic leukemia. Blood Cells Mol Dis. 2013;50(1):39-40
  44. Protosaltis NJ, Liang W, Nudleman E, Ferrara N. Interleukin-22 promotes tumor angiogenesis. Angiogenesis.